

# Fundamentals of the objectiveFIELD® Analyzer (OFA®)

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## Why do we need a new perimetric device?

The answer is partly that while Standard Automated Perimetry (**SAP**) remains the cornerstone of visual field testing, it has limitations that are perhaps not widely appreciated. Additionally, a different type of perimeter could provide extra statistically independent types of information that when combined could improve diagnostic and prognostic power. A more user-friendly device would also be welcome.

## SAP limitations

SAP is a behavioural button-press test in which its stimuli are presented one-at-a-time to find a proxy for true sensitivity, a light threshold, at each location. Because the process is laborious and uncomfortable, participants often find it unpleasant,<sup>1</sup> and *it poses particular challenges for younger, or older persons, or physically frail individuals*.

Perhaps the biggest problem with SAP is that it suffers from poor reproducibility. This originates in part from the under-sampling of abrupt changes in sensitivity associated with visual field defects by SAP's typical Goldmann Size-3 stimuli, which are only 0.43 deg across. Even the relatively dense SAP 10-2 array of stimuli (**Fig. 1A**) leaves 96.4% of the macula untested, while the more commonly used and spatially broader SAP 24-2 pattern *misses* 99.6%. We have shown that SAP's poor sampling of the retina contributes greatly to its poor reproducibility<sup>2, 3</sup> Dysfunctional retinal neurons also contribute, but their contribution is about 12-times smaller.<sup>4</sup> Poor reproducibility impairs our ability to detect clinical changes over time, i.e. it affects *prognostic* power. Even a rapidly declining glaucoma patient needs to have 6 SAP tests over a 2-year period to have even an 80% chance of that decline being detected.<sup>5</sup> *In a busy clinical setting, this is rarely feasible*. By contrast, the *objectiveFIELD Analyser* (**OFA**, **Fig. 2A**), has better reproducibility, partly due to its larger stimuli (*cf.* the white dots in Fig. 1A with the yellow regions in **Fig. 1B**), and is faster than SAP.<sup>6</sup> The macular OFA stimulus ensemble of Fig. 1B makes comparisons with OCT retinal thickness data easier. OFA also has widefield tests. OFA measures physiological responses directly from the test subject through their pupils: users need only fixate their vision on a centrally located cross. So, it uses no button presses.

Another weakness of SAP is that the sensitivity losses it reports follow early retinal cell losses poorly. That is, changes in SAP sensitivity are a nonlinear function of ganglion cell loss.<sup>7</sup> As the eye damage progresses from minimal to quite severe cell loss, the slope of the function changes very little – the functional changes reflected by SAP are not truly representative of the severity of cell loss. Only after substantial damage has occurred do increases in damage

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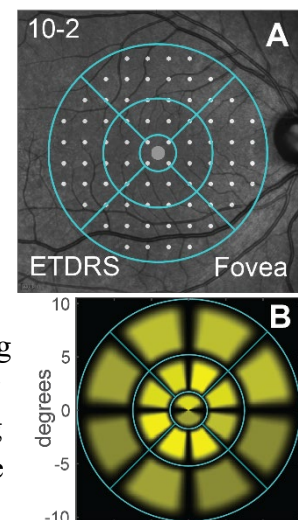
reported by SAP then change rapidly. By contrast, the sensitivity changes measured by OFA follow the decline in cell numbers closely (i.e. linearly), in principle allowing OFA to report damage earlier.<sup>8</sup> Plotting SAP mean defects (MDs) on OFA MDs in the same subjects shows this nonlinearity convincingly.<sup>6</sup>

A further issue with SAP is that the small size of Goldmann Size-3 stimuli means that the retinal contrast gain control system of magnocellular retinal ganglion cells (M-cells) that operates for natural stimuli is not engaged. When operating normally that system can change the responsiveness (gain) of M-cells by an order of magnitude in tens of milliseconds. To control that accurately, it uses a rapidly computed spatial average of contrasts over an area substantially larger than the M-cell inhibitory surround.<sup>9</sup> Macaque studies show that retinal gain control does not therefore operate for stimuli  $\leq 0.5$  deg across,<sup>10</sup> i.e. the diameter of Goldmann Size-3 stimuli. The larger, high contrast stimuli of OFA (e.g. Fig. 1B) thus test the visual system as it was designed to operate: in a world of dynamically changing spatially extensive stimuli. A corollary of that is that new information about the effects of glaucoma and other diseases upon that dynamic system may be revealed.

## OFA Basics

OFA uses multifocal stimuli where the measured physiological response is the *relative* change in pupil diameter. Using relative change in pupil size is very commonly done,<sup>11</sup> partially because the pupil response dynamics are independent of size.<sup>12</sup> This also means that, in most cases, drugs that mildly alter pupil size, and at least one pupil is not stationary or quite irregular, are not an issue. Local iris defects caused by things like cataract surgery are generally not a problem. Age effects are also reduced. Ptosis presents a possible issue for OFA, but it doesn't track superior pupil size in larger pupils. During testing, independent stimuli are presented to the two eyes concurrently (halving test duration) and both pupils are recorded. The resulting direct and consensual responses are combined according to the measured signal to noise ratios, thus one poorly

performing pupil is not a large issue. Pupil responses are sometimes thought to be sub-cortical, but this is only true for the extremely slow responses of the melanopsin containing retinal ganglion cells,<sup>13</sup> which regulate pupil size in bright light. Input to the pupil system from many cortical areas is well established.<sup>14</sup> We have provided evidence that the transient onset stimuli of OFA drive a cortical pathway,<sup>15</sup> probably through V2.<sup>16</sup> This appears to be an offshoot from the neural wiring supporting the accommodative triad. That system needs a rapid estimate of range to objects as is provided by stereopsis, which is computed in the cortex.<sup>17</sup>



**Fig. 1 A)** The macula as delineated by the cyan **ETDRS\*** grid. The central grey dot is the foveola. The small white dots are the tiny 10-2 SAP stimuli, which fail to test 96.4% of the macula. **B)** The OFA macular 10-EDTRS stimulus ensemble shown relative to the ETDRS grid. OFA also has wide-field stimuli testing  $\pm 30$  degrees of the field. \**Early Treatment Diabetic Retinopathy Study*.

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OFA methods have been improving over time. Three innovations have been new stimulus methods each of which improved signal-to-noise ratios by 40%. These included: luminance balancing,<sup>18</sup> and clustered volleys.<sup>19</sup> Most recently, methods for fitting pupillary gain dynamics have done things like permit widefield<sup>20</sup> and macular methods<sup>21</sup> that test both eyes in under 90 seconds. With a few exceptions, OFA papers published since 2020 use the latest test methods (N=13). OFA has been demonstrated to be safe for individuals with migraine<sup>22</sup> and epilepsy.<sup>23</sup>

### Dual-axis *versus* Half-axis testing

Another issue with SAP is that it is a *Half-axis* device because it only reports negative changes in sensitivity (**Fig. 2B**). It does not capture *increases* in sensitivity that may be indicative of phenomena such as glutamate excitotoxicity.<sup>24</sup> By contrast OFA is a *Dual-axis* device reporting increases and decreases in both sensitivity and response delays at every tested region. As will be described below, response delays often include information about damage that is independent of sensitivity, i.e. which may report on independent aspects of disease.

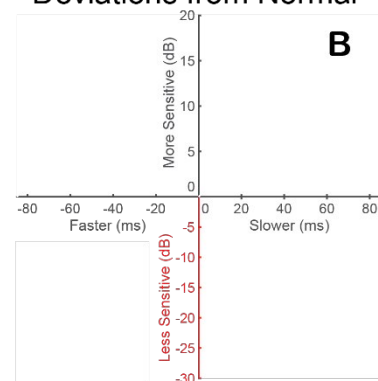
Hypersensitivity is an interesting example. We have shown that higher than normal sensitivities (hypersensitivity) are associated with early-stage diabetic retinal disease (DRD), e.g.<sup>25, 26</sup>, providing better diagnostic power than SAP.<sup>26</sup> Hypersensitivities can predict which AMD eyes commencing anti-VEGF therapy will respond well, both initially<sup>27</sup> and over 15 months.<sup>28</sup> SAP testing using larger stimuli, that would engage retinal gain control, have been reported to show hypersensitive regions in glaucoma. Casson and Johnson referred to these areas as “high-sensitivity defects”,<sup>29</sup> a nice succinct term. These could decay to regions of decreased sensitivity (“low-sensitivity defects”) over 2 to 3 years.<sup>29</sup> Notice that for SAP to quantify hypersensitivity requires people to detect miniscule changes in contrast while on OFA hypersensitivity is an easy-to-measure larger than normal response.

Longer response delays are characteristic of later-stage DRD, and oedema related-changes in retinal thickness over 2 years, while SAP reports no change in the same subjects.<sup>30</sup>

Our 5-year study of glaucoma showed that some eyes progress on sensitivity but not delay, or *vice versa*.<sup>31</sup> That phenomenon has been partially confirmed by others using an independent method: saccadic perimetry.<sup>32</sup> As in OFA, regions showing large response delays can occur where (nonlinear) SAP reports normal sensitivity. Others have reported increased saccadic delay and fewer express saccades in glaucoma.<sup>33</sup> Delays and sensitivities are differently correlated with AMD severity and Macular Pigment Optical Density,<sup>34</sup> indicating independent measures of the disease. OFA delays differentiate focal and generalised epilepsy,<sup>23</sup> provide high diagnostic power in multiple



Dual-axis vs. Half-axis  
Deviations from Normal



**Fig. 2** A) An OFA in the clinic showing a test subject and the operator view of the videoed pupils. B) The difference between a Half-axis measuring device like SAP (red axis), and a Dual-axis device like OFA providing a broader data set.

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sclerosis (MS),<sup>20</sup> and predict which MS subjects will change to progressive disease over 10 years.<sup>35</sup> Delays and sensitivities are differently affected at different field locations following mild concussion.<sup>36</sup> Overall, we have shown that measuring both sensitivity and delays can add significantly to diagnostic and prognostic power.

For example, we compared the diagnostic power of 44 different functional and structural tests from 23 studies attempting to discriminate eyes of normal controls from diabetic persons with no retinopathy (i.e. very early-stage DRD). The result was that OFA has by far the best performance.<sup>37</sup>

As OFA uses pupil responses, the issue of iris neuropathy arises. Iris-specific issues will affect all regions equally, either sensitivities or delays. Therefore, like the effects of cataract in SAP, the OFA pattern deviations will be unaffected by such global biases. Thus, pattern deviations will resolve true regional delay defects. We have quantified localised delay defects in early-stage DR independent of other factors.<sup>26</sup>

### Long regional delays masquerading as SAP sensitivity loss?

The response delays observed with OFA can be upwards of 600 ms. This is comparable to the delays reported for saccadic perimetry in glaucoma.<sup>32</sup> Two studies have examined response delays in glaucoma patients when they were given more time to respond in SAP-like testing.<sup>38, 39</sup> In one study 25% of the glaucoma subjects had a mean delay of > 1 second.<sup>38</sup> Such delays could mean that in normal SAP testing that stimuli are missed, which are then interpreted as sensitivity loss. Another study was interested in performance at various limens of the contrast-response function for SAP-like stimuli.<sup>39</sup> Damaged areas of the field had mean delays of 996.3 ms for the 0 dB stimulus. In damaged areas six out of ten persons with glaucoma registered no response at all within the 2 s of allowed time. These studies suggest it is possible that long response delays could masquerade as SAP sensitivity loss. We followed glaucoma subjects over 5 years and demonstrated that sensitivity and delay losses can progress quite independently over time and across the fields.<sup>31</sup> Thus, a false SAP defect, due to a long response delay, could easily occur at a different location to true sensitivity loss. Independent evolution of OFA sensitivity and delay defects is in line with results from saccadic perimetry, and we have reported such results for OFA in MS,<sup>35</sup> DR,<sup>26, 30, 40</sup> and AMD.<sup>28</sup>

Returning to delayed responses using SAP-like stimuli, an interesting aspect of one of those studies was that persons with more acute diseases, AION and Stroke, had shorter response delays than the glaucoma patients.<sup>38</sup> That might suggest that some regional response delays might be due to subsequent degeneration of the cortex and optic radiations that has been reported in glaucoma.<sup>41, 42</sup> That appears to mirror demonstrations of trans-neuronal retrograde degeneration of macular retinal ganglion cells (RGCs) following striate cortical ablation in macaque monkeys.<sup>43</sup> In such studies RGC loss increases over periods between 1 and 9 years.<sup>44</sup> Both visual search<sup>45</sup> and reading speeds<sup>46</sup> are slower in persons with glaucoma. Comparing those data with OFA sensitivity and delay data would be interesting. Repeating those comparisons in persons with recent acute glaucoma (e.g. uveitic) and chronic glaucoma over several years could also be informative.

## Summary

The 39 [OFA publications](#) to date include studies of 8 ophthalmic and brain diseases. The dual-axis OFA results appear to provide new, and statistically independent, information, improving diagnosis and prognosis. Its two high-resolution tests provide four 30-2 like reports in 8 minutes.<sup>6</sup> Its three lower-resolution 90-second tests are particularly useful for children<sup>47</sup> and infirm persons.<sup>48</sup> Both types of tests come in macular and wide-field versions. The stimuli of the fast macular test<sup>21</sup> match the test regions of the ETDRS grid used by OCTs (Fig. 1B) to report retinal thickness data, simplifying structure/function comparisons.

## References

1. Glen FC, Baker H, Crabb DP. A qualitative investigation into patients' views on visual field testing for glaucoma monitoring. *BMJ Open* 2014;4:e003996. [10.1136/bmjopen-2013-003996](#)
2. Maddess T. The influence of sampling errors on test-retest variability in perimetry. *Invest Ophthalmol Vis Sci* 2011;52:1014-1022. [10.1167/iovs.10-6014](#)
3. Maddess T. Modelling the relative influence of fixation and sampling errors on test-retest-variability in perimetry. *Graefes Archive Ophthalmol* 2014;252:1611–1619. [10.1007/s00417-014-2751-y](#)
4. Numata T, Maddess T, Matsumoto C, Okuyama S, et al. Exploring test-retest variability using high-resolution perimetry. *Trans Vis Sci Tech* 2017;6:1-9. [10.1167/tvst.6.5.8](#)
5. Chauhan BC, Garway-Heath DF, Goni FJ, Rossetti L, et al. Practical recommendations for measuring rates of visual field change in glaucoma. *Br J Ophthalmol* 2008;92:569-573. [10.1136/bjo.2007.135012](#)
6. Maddess T, Carle CF, Kolic M, Essex RW, et al. Diagnostic power and reproducibility of objective perimetry in glaucoma. *J Glaucoma* 2024;32:940-950. [10.1097/IJG.0000000000002485](#)
7. Hood DC, Anderson SC, Wall M, Kardon RH. Structure versus function in glaucoma: an application of a linear model. *Invest Ophthalmol Vis Sci* 2007;48:3662-3668. [10.1167/iovs.06-1401](#)
8. Carle CF, Chain AYH, Kolic M, Maddess T. The structure-function relationship between multifocal pupil perimetry and retinal nerve fibre layer in glaucoma. *BMC Ophthalmology* 2024;24:1-11. [10.1186/s12886-024-03402-z](#)
9. Hochstein S, Shapley RM. Linear and nonlinear spatial subunits in Y cat retinal ganglion cells. *J Physiol* 1976;262:265-284. [10.1113/jphysiol.1976.sp011595](#)
10. Benardete EA, Kaplan E. The dynamics of primate M retinal ganglion cells. *Vis Neurosci* 1999;16:355-368. [10.1017/s0952523899162151](#)
11. Bremner FD. Pupillometric evaluation of the dynamics of the pupillary response to a brief light stimulus in healthy subjects. *Invest Ophthalmol Vis Sci* 2012;53:7343-7347. [10.1167/iovs.12-10881](#)
12. Maddess T. Pupil dynamics and response amplitude: only size matters. *Invest Ophthalmol Vis Sci* 2012;53: [10.1167/iovs.12-11105](#)



13. Dacey DM, Liao HW, Peterson BB, Robinson FR, et al. Melanopsin-expressing ganglion cells in primate retina signal colour and irradiance and project to the LGN. *Nature* 2005;433:749-754. [10.1038/nature03387](https://doi.org/10.1038/nature03387)
14. Gamlin PD. The pretectum: connections and oculomotor-related roles. *Prog Brain Res* 2006;151:379-405. [10.1016/S0079-6123\(05\)51012-4](https://doi.org/10.1016/S0079-6123(05)51012-4)
15. Carle CF, James AC, Kolic M, Essex RW, et al. Blue Multifocal Pupillographic Objective Perimetry in Glaucoma. *Invest Ophthalmol Vis Sci* 2015;56:6394-6403. [10.1167/iovs.14-16029](https://doi.org/10.1167/iovs.14-16029)
16. Sabeti F, James AC, Carle CF, Essex RW, et al. Comparing multifocal pupillographic objective perimetry (mfPOP) and multifocal visual evoked potentials (mfVEP) in retinal diseases. *Sci Report* 2017;7:45847. [10.1038/srep45847](https://doi.org/10.1038/srep45847)
17. Suryakumar R, Allison R. Accommodation and pupil responses to random-dot stereograms. *J Optom* 2016;9:40-46. [10.1016/j.optom.2015.03.002](https://doi.org/10.1016/j.optom.2015.03.002)
18. Carle CF, James AC, Kolic M, Essex RW, et al. Luminance and colour variant pupil perimetry in glaucoma. *Clin Exp Ophthalmol* 2014;42:815-824. [10.1111/ceo.12346](https://doi.org/10.1111/ceo.12346)
19. Carle CF, James AC, Sabeti F, Kolic M, et al. Clustered Volleys stimulus presentation for multifocal objective perimetry. *Trans Vis Sci Tech* 2022;11(2):1-10. [10.1167/tvst.11.2.5](https://doi.org/10.1167/tvst.11.2.5)
20. Maddess T, van Kleef JP, Rohan EMF, Carle CF, et al. Rapid, non-contact multifocal visual assessment in multiple sclerosis. *Neurol Sci* 2022;43:1-7. [DOI: 10.1007/s10072-022-06387-z](https://doi.org/10.1007/s10072-022-06387-z)
21. Rai BB, Sabeti F, Carle CF, Rohan EMF, et al. Rapid objective testing of visual function matched to the ETDRS-grid, and its diagnostic power in AMD. *Ophthalmol Sci* 2022;2:1-9. [10.1016/j.xops.2022.100143](https://doi.org/10.1016/j.xops.2022.100143)
22. Ali EN, Carle CF, Lueck CJ, Kolic M, et al. Assessing migraine patients with multifocal pupillographic objective perimetry. *BMC Neurol* 2021;21:1-12. [10.1186/s12883-021-02239-z](https://doi.org/10.1186/s12883-021-02239-z)
23. Ali EN, Lueck CJ, Carle CF, Martin KL, et al. Response characteristics of objective perimetry in persons living with epilepsy. *J Neurol Sci* 2022;436:120237. [10.1016/j.jns.2022.120237](https://doi.org/10.1016/j.jns.2022.120237)
24. Feng L, Dai S, Zhang C, Zhang W, et al. Ripa-56 protects retinal ganglion cells in glutamate-induced retinal excitotoxic model of glaucoma. *Sci Rep* 2024;14:3834. [10.1038/s41598-024-54075-z](https://doi.org/10.1038/s41598-024-54075-z)
25. Bell A, James AC, Kolic M, Essex RW, et al. Dichoptic multifocal pupillography reveals afferent visual field defects in early type 2 diabetes. *Invest Ophthalmol Vis Sci* 2010;51:602-608. [10.1167/iovs.09-3659](https://doi.org/10.1167/iovs.09-3659)
26. Sabeti F, van Kleef JP, Iyer RM, Carle CF, et al. Discriminating early-stage diabetic retinopathy with subjective and objective perimetry. *Frontiers Endo* 2024;14:1-12. [10.3389/fendo.2023.1333826](https://doi.org/10.3389/fendo.2023.1333826)
27. Sabeti F, Maddess T, Essex RW, James AC. Multifocal pupillography identifies ranibizumab induced changes in retinal function for exudative age-related macular degeneration. *Invest Ophthalmol Vis Sci* 2012;53:253-260. [10.1167/iovs.11-8004](https://doi.org/10.1167/iovs.11-8004)

28. Rai BB, Essex RW, Sabeti F, Maddess T, et al. An objective perimetry study of central versus. peripheral sensitivities and delays in age-related macular degeneration. *Trans Vis Sci Tech* 2021;10:1-14. [10.1167/tvst.10.14.24](#)
29. Casson EJ, Johnson CA. Flicker perimetry in ocular hypertensives: evidence for high- and low-sensitivity defects. *Noninvasive Assessment of the Visual System, Technical Digest Series* 1993;NMC.4:304-307. [10.1364/NAVS.1993.NMC4](#)
30. Sabeti F, Rai BB, van Kleef JP, Carle CF, et al. Objective perimetry identifies functional progression and recovery in mild diabetic macular oedema. *PLoS One* 2023;18: e0287319. [10.1371/journal.pone.0287319](#)
31. Maddess T, Carle CF, Kolic M, Sarac O, et al. Independence across space and time of sensitivity and delay visual field measures in glaucoma. . *IPS* 2024. [www.perimetry.org/abstracts/](http://www.perimetry.org/abstracts/)
32. Thepass G, Lemij HG, Vermeer KA, van der Steen J, et al. Slowed saccadic reaction times in seemingly normal parts of glaucomatous visual fields. *Front Med (Lausanne)* 2021;8:679297. [10.3389/fmed.2021.679297](#)
33. Kanjee R, Yucel YH, Steinbach MJ, Gonzalez EG, et al. Delayed saccadic eye movements in glaucoma. *Eye Brain* 2012;4:63-68. [10.2147/EB.S38467](#)
34. Rai BB, Sabeti F, van Kleef JP, Carle CF, et al. Comparing 2-dimensional macular pigment optical density and macular function measured by objective perimetry in early to late stage AMD. *Graef's Archive Clin Exp Ophthalmol* 2024;262:1-11. [10.1007/s00417-024-06437-6](#)
35. Maddess T, Carle CF, Rohan EMF, Baird-Gunning J, et al. Objective perimetry and progression of multiple sclerosis. *eNeurol Sci* 2022;100430:1-6. [10.1016/j.ensci.2022.100430](#)
36. Sabeti F, Carle CF, Jaros RK, Rohan EMF, et al. Objective perimetry in sporting-related mild traumatic brain injury. *Ophthalmology* 2019;126:1053-1055. [10.1016/j.ophtha.2019.01.026](#)
37. Rai BB, van Kleef JP, Sabeti F, Vlieger R, et al. Early diabetic eye damage: comparing detection methods using diagnostic power. *Survey Ophthal* 2024;69:24-33. [10.1016/j.survophthal.2023.09.002](#)
38. Nowomiejska K, Vonthein R, Paetzold J, Zagorski Z, et al. Reaction time during semi-automated kinetic perimetry (SKP) in patients with advanced visual field loss. *Acta Ophthalmol* 2010;88:65-69. [10.1111/j.1755-3768.2008.01407.x](#)
39. Wall M, Maw RJ, Stanek KE, Chauhan BC. The psychometric function and reaction times of automated perimetry in normal and abnormal areas of the visual field in patients with glaucoma. *Invest Ophthalmol Vis Sci* 1996;37:878-885. [iovs.arvojournals.org/article.aspx?articleid=2180501](http://iovs.arvojournals.org/article.aspx?articleid=2180501)
40. Rai BB, Maddess T, Carle CF, Rohan EMF, et al. Comparing objective perimetry, matrix perimetry, and regional retinal thickness in early diabetic macular oedema. *Trans Vis Sci Tech* 2021;10:1-12. [10.1167/tvst.10.13.32](#)
41. Gupta N, Ang LC, Noel de Tilly L, Bidaisee L, et al. Human glaucoma and neural degeneration in intracranial optic nerve, lateral geniculate nucleus, and visual cortex. *Br J Ophthalmol* 2006;90:674-678. [10.1136/bjo.2005.086769](#)

42. Zhang QJ, Wang D, Bai ZL, Ren BC, et al. Diffusion tensor imaging of optic nerve and optic radiation in primary chronic angle-closure glaucoma using 3T magnetic resonance imaging. *Int J Ophthalmol* 2015;8:975-979. [10.3980/j.issn.2222-3959.2015.05.22](https://doi.org/10.3980/j.issn.2222-3959.2015.05.22)
43. Johnson H, Cowey A. Transneuronal retrograde degeneration of retinal ganglion cells following restricted lesions of striate cortex in the monkey. *Exp Brain Res* 2000;132:269-275. [10.1007/s002210000384](https://doi.org/10.1007/s002210000384)
44. Cowey A, Stoerig P, Williams C. Variance in transneuronal retrograde ganglion cell degeneration in monkeys after removal of striate cortex: effects of size of the cortical lesion. *Vision Res* 1999;39:3642-3652. [10.1016/s0042-6989\(99\)00097-8](https://doi.org/10.1016/s0042-6989(99)00097-8)
45. Sun MJ, Rubin GS, Akpek EK, Ramulu PY. Impact of glaucoma and dry eye on text-based searching. *Transl Vis Sci Technol* 2017;6:24. [10.1167/tvst.6.3.24](https://doi.org/10.1167/tvst.6.3.24)
46. Rolle T, Dallorto L, Cafasso R, Mazzocca R, et al. Reading ability in primary open-angle glaucoma: evaluation with radner reading charts. *Optom Vis Sci* 2019;96:55-61. [10.1097/OPX.0000000000001319](https://doi.org/10.1097/OPX.0000000000001319)
47. Maddess T, Rohan EMF, Rai BB, Carle CF, et al. Diagnostic power of rapid objective perimetry in young people with Type 1 Diabetes. *IOVS-ARVO* 2023;64:e2666. [iovs.arvojournals.org/article.aspx?articleid=2790300](https://iovs.arvojournals.org/article.aspx?articleid=2790300)
48. Maddess T, Rohan EMF, Kulh MA, van Kleef JP, et al. Diagnostic power of rapid multifocal objective perimetry in Alzheimer's. *IOVS-ARVO* 2024;65:e83. [iovs.arvojournals.org/article.aspx?articleid=2794574](https://iovs.arvojournals.org/article.aspx?articleid=2794574)